# **POLB knockout is synthetic lethal with PARP MARIANCE TANGO** therapeutics<sup>™</sup> inhibition leading to complete and durable responses in BRCA-mutant tumor xenografts



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ABSTRACT	POLB knockout enhances PARP inhibitor-induced growth inhibition in BRCA2 mutant cells	Knockout of POLB in BRCA1 mutant tumors enhances response to PARP inhibitor treatment
Despite the clinical benefit of PARP1/2 inhibitors (PARPi), which are FDA-approved for the treatment of certain BRCA-mutant cancers, many patients achieve incomplete disease control and develop progressive disease. Motivated by this clinical need, we utilized our CRISPR target discovery screening platform to identify novel targets that synergize with PARP inhibitor treatment. By conducting parallel screens in both BRCA-mutant and wildtype cells, we identified DNA polymerase beta (POLB) as a novel target that - when combined with PARPi - selectively kills BRCA-mutant lines while sparing normal cells. POLB knockout and cDNA rescue experiments using both BRCA1 and BRCA2-mutant isogenic cell lines further demonstrated that the catalytic activity of POLB is required for synthetic lethality with PARPi. Most strikingly, POLB knockout combined with sub-therapeutic doses of PARPi, led to profound tumor regression and prevented in vivo tumor regrowth, even after cessation of	A B BRCA2 WT NTC POLB KO POLB O POLB O POLB O POLB O POLB O POLB O POLB O	<pre>2000 - NTC (Vehicle) - NTC (Niraparib) - NTC (Niraparib) - POLB KO (Vehicle) - ▲ POLB KO (Niraparib)</pre>

drug treatment. Mechanistically, POLB knockout is associated with increased single and double strand DNA breaks, accumulation of poly-ADP-ribose polymers, cell cycle arrest, and apoptosis. Together, these results suggest that POLB inhibitors in combination with PARPi have the potential to drive deep and durable responses providing a novel therapeutic option for cancer patients with BRCA1/2-mutations.

Abstract #10







Figure 7: Loss of POLB in BRCA1 mutant cells enhances PARP inhibitor-induced tumor growth inhibition. Xenograft tumors generated from Nontargeting control and POLB knockout MDA-MB-436 cells treated with either vehicle or 15mpk Niraparib for 28 days, PO, QD dosing.

### Complete and durable tumor growth inhibition with POLB KO + PARP inhibitor treatment



Figure 1: CRISPR-based drug anchor screens in the presence or absence of Olaparib. (A) Schematic of CRISPR drug anchor screens performed with +/- Olaparib in 2 BRCA1 mutant cell lines and DLD1 isogenic cell lines. sgRNA enrichment/depletion analysis represented in volcano plots. (B) MDA-MB-436. (C) A549. (D) DLD1 BRCA2 null (-/-). (E) DLD1 BRCA1/2 WT (+/+).

## POLB KO



Figure 4: POLB knockout enhances PARPi-induced growth inhibition in BRCA2 mutant cells. POLB knockout and non-targeting sgRNA control cells were generated using DLD1 isogenic cell lines. (A, B) Western blot confirming POLB knockout. (C) 7-day viability assay with Niraparib. (D) 7-day viability assay with Olaparib. (E) 14-day colony formation assays with Niraparib. (F) 14-day colony formation assays with Olaparib.

### POLB sgRNAs drop out with PARP inhibitor treatment in a time, dose, and **BRCA-dependent manner**



Figure 2: Dropout validation screen of seven independent POLB sgRNAs in DLD1 isogenic cell lines with PARP inhibitor treatment: Fold change (FC) in POLB sgRNAs compared to DMSO control group.

POLB knockout enhances PARP inhibitor response in BRCA1 mutant cells

#### Inhibition of polymerase and lyase activities of POLB are required for synthetic lethal activity with PARP inhibitor



Days post first treatment

Figure 8: Loss of POLB leads to complete and durable response with PARP inhibitor treatment. (A) Xenograft tumors generated from DLD1 BRCA2 null POLB knockout cells treated with either vehicle or Niraparib at 30mpk, PO, QD dosing. (B) Tumors generated from DLD1 BRCA2 null POLB knockout cells from individual animals either treated with vehicle or niraparib post cessation of treatment at 28 days.

## SUMMARY

• Olaparib drug anchor screens identify POLB as a synthetic lethal target in BRCA1/2 mutant cancers

 Genetic validation confirms strong synergy with PARP inhibitor in vitro and in vivo at clinically relevant doses, including complete and durable response after cessation of niraparib treatment

• Inhibition of polymerase and lyase activity of POLB is required for the synthetic lethal interaction with PARP inhibitors based on cDNA rescue studies

• Knockout of POLB with PARP inhibitor treatment induces single-strand breaks, leading to doublestrand breaks, and enhanced cell cycle arrest and apoptosis supporting a synthetic lethal model

BRCA1/2 WT	Loss of functional BRCA1/2	Loss of functional BRCA1/2	

Figure 5: Dual loss of Polymerase and Lyase activities of POLB are required for synthetic lethal activity with PARPi: (A) Crystal structure of POLB showing catalytic residues of polymerase and lyase activities (Ref 1). (B) Immunoblotting analysis confirming POLB WT cDNA or mutant cDNA overexpression (C) 7-day viability assay with Niraparib.







ACKNOWLEDGEMENTS

The authors gratefully acknowledge the generous contributions from teams at Biortus, ChemPartner, WuXi AppTec, and intoDNA

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Figure 3: POLB knockout enhances PARP inhibitor response in BRCA1 mutant cell lines. POLB knockout and non-targeting control (NTC) cell lines were generated in BRCA1 mutant MDAMB436 cells (top panel) and SUM149PT cells (bottom panel). (A, D) Immunoblotting confirming POLB knockout. (B, E) 7-day CTG viability assay with Niraparib. (C, F) 7-day CTG viability assay with Olaparib.

Figure 6: Quantification of single-strand breaks in response to POLB knockout (A) sSTRIDE analysis to assess single-strand breaks (Blue, nuclear stain; Pink, single strand breaks) in DLD1 BRCA2 mutant cells. sSTRIDE technology is described in Ref 2. (B) Graph representing the quantification of single-strand breaks in more than 1000 cells per sample.