



Abstract #3363

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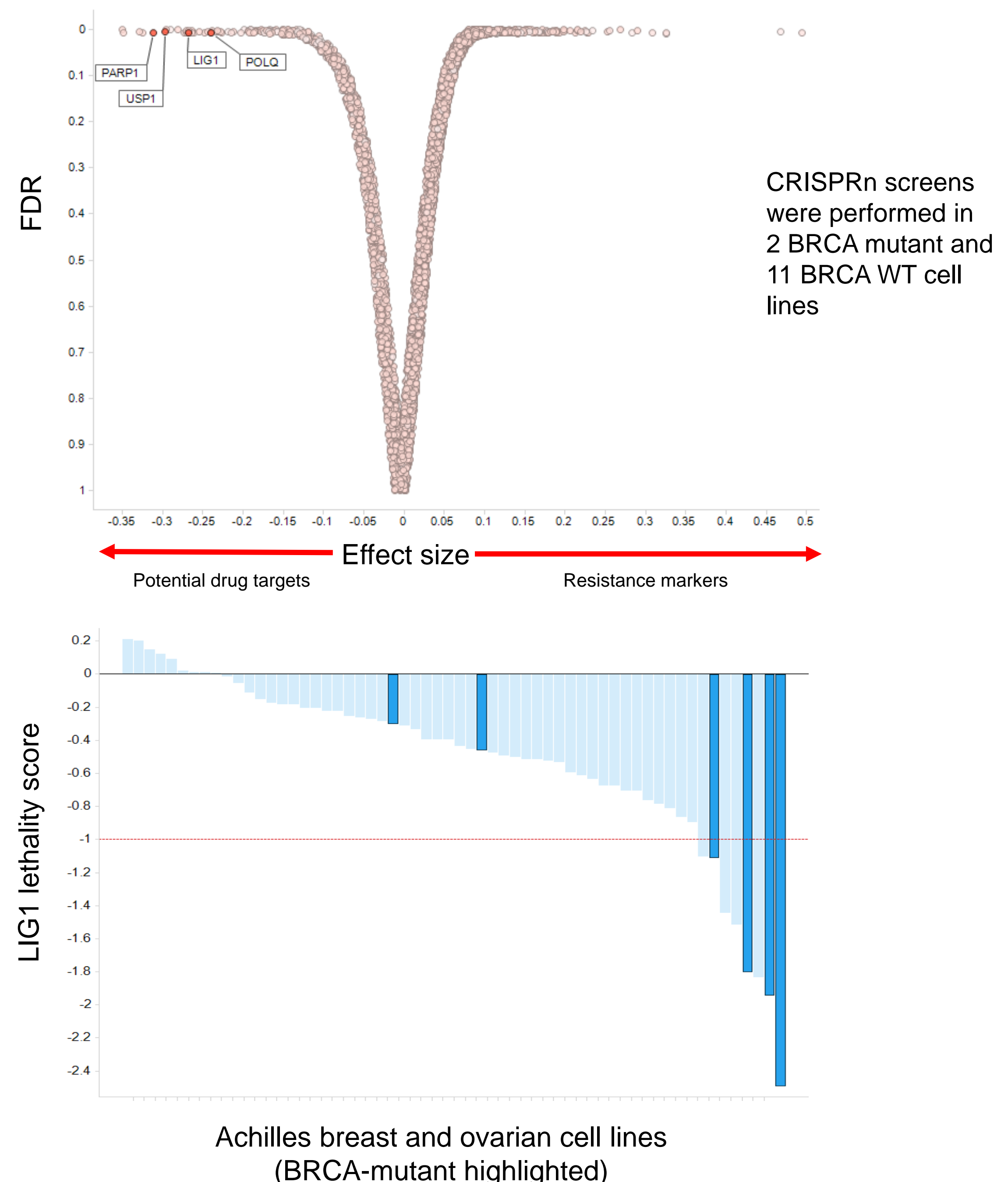
## Introduction

Women harboring a mutation in the *BRCA1* gene have a >7X increased risk of developing breast cancer and >35X increased risk of developing ovarian cancer in their lifetime compared to women lacking such mutations. While PARP1/2 inhibitors are efficacious in the *BRCA* mutant setting, this efficacy is limited and resistance develops readily. Additional therapeutic targets beyond PARP1/2 are needed to treat cancer patients with *BRCA* mutations.

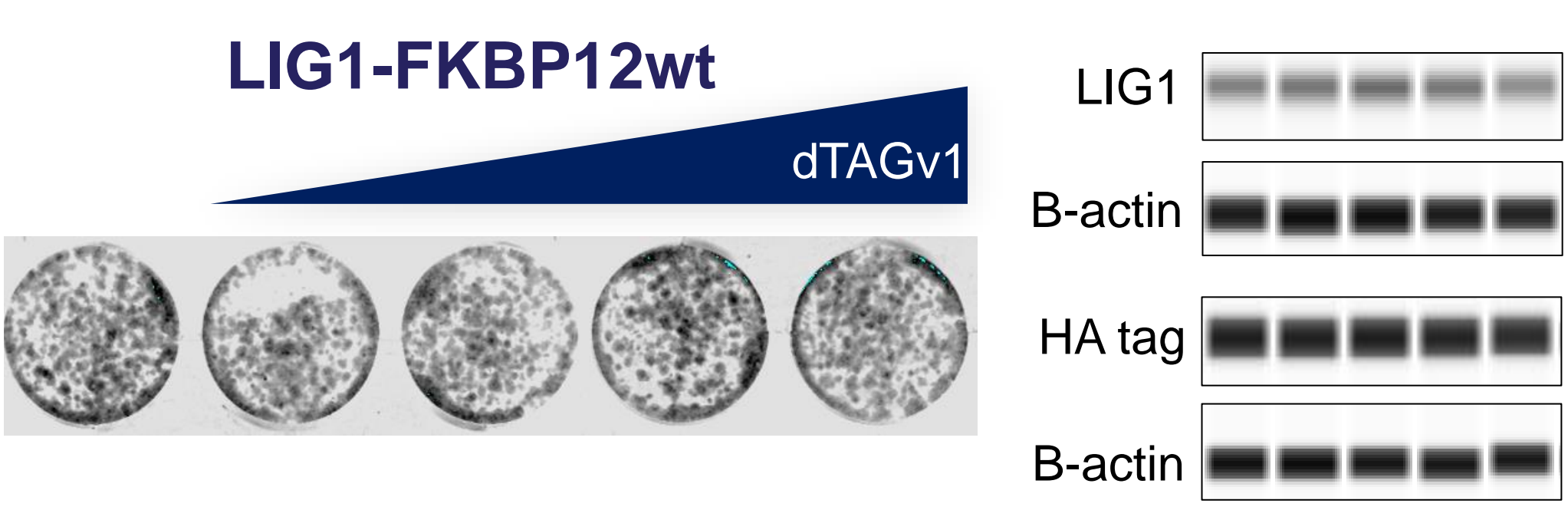
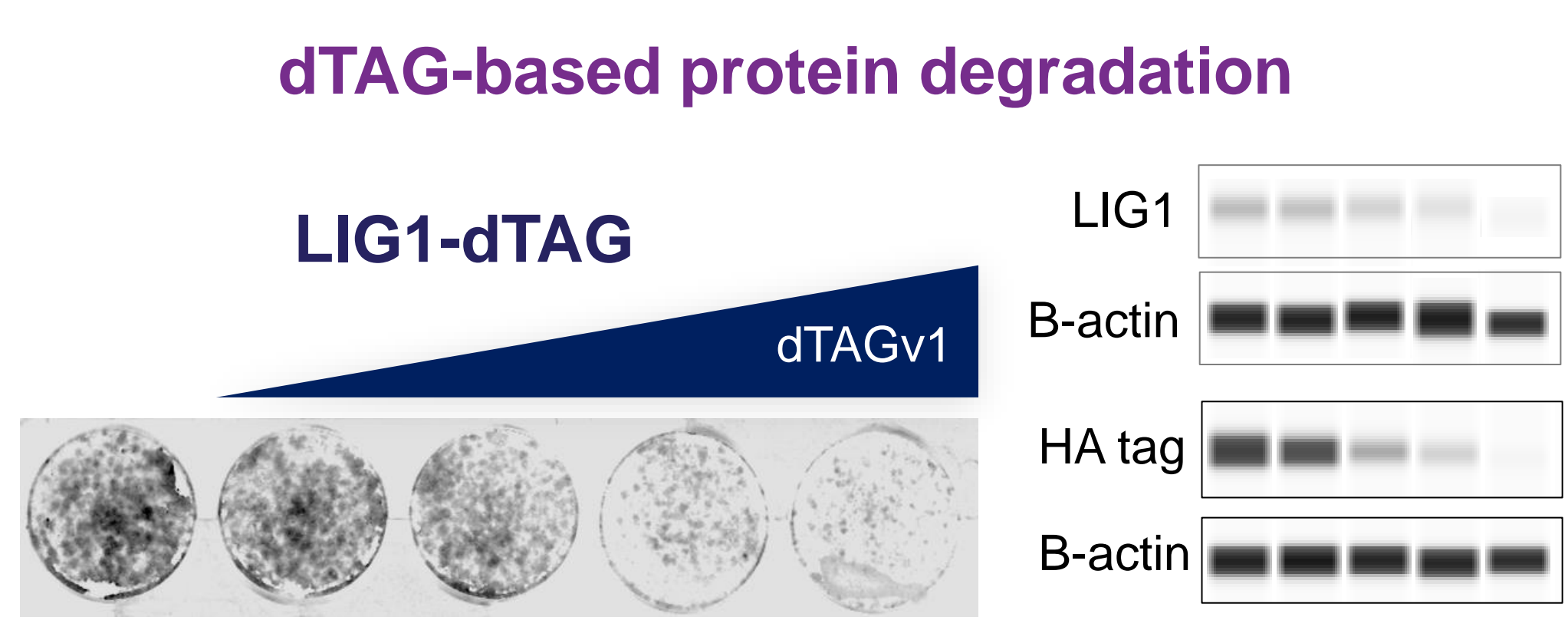
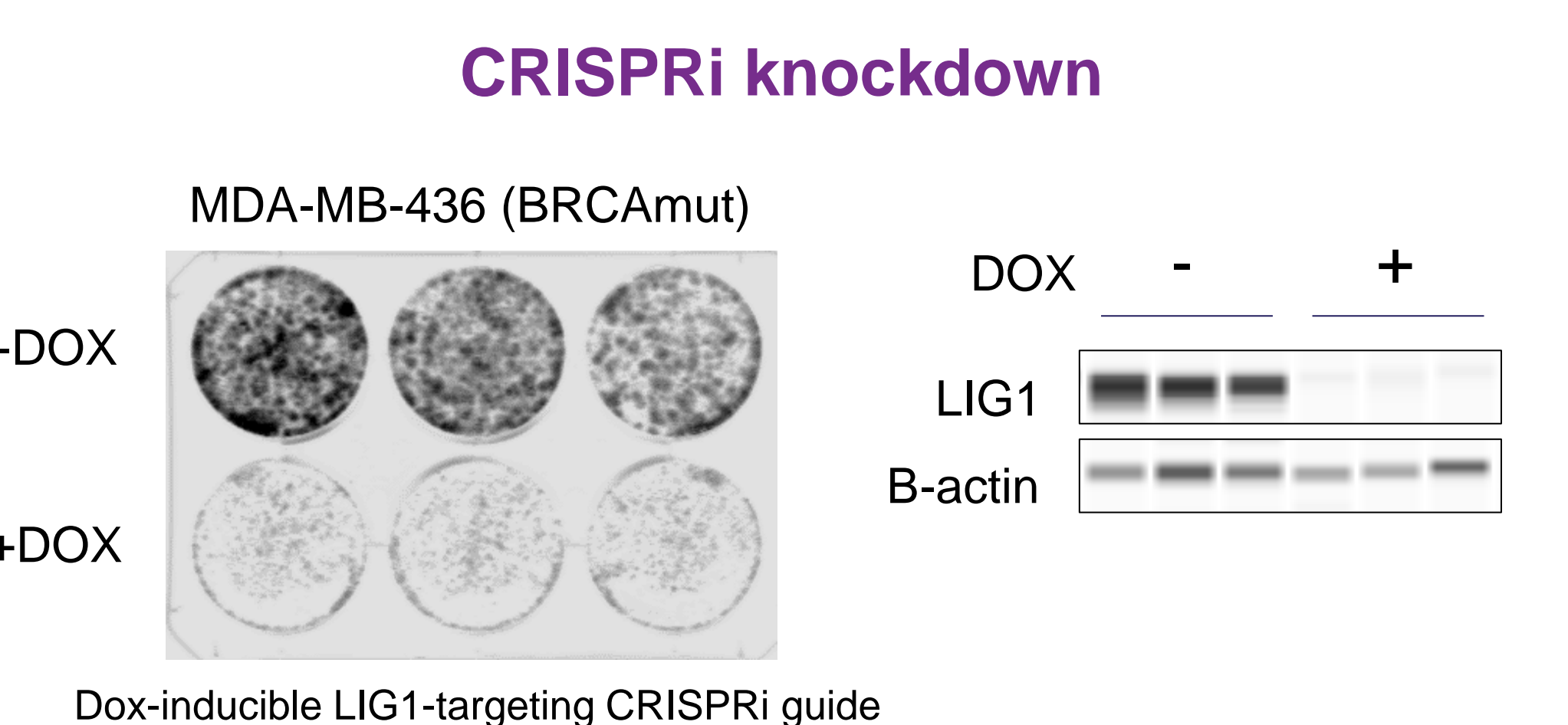
We performed CRISPR screens in 11 *BRCA1/2* wild-type and 2 *BRCA1* mutant cancer cell lines to identify targets that are synthetic lethal with *BRCA1* mutations. In addition to USP1, PARP1 and POLQ, we identified the DNA ligase gene *LIG1* as a novel target that when knocked out, kills *BRCA1* mutant cells selectively. Internal analysis of *BRCA* mutations in breast and ovarian cancer cell lines in the Project Achilles database further validated the hyperdependence of *BRCA1* mutant cells on *LIG1*. Single-gene perturbation of *LIG1* using CRISPRn, CRISPRi, and RNAi confirmed the lethal effect of *LIG1* inactivation in *BRCA1* mutant cell lines, but not *BRCA1/2* wild-type cell lines. This loss of viability could be rescued by complementing with an exogenous wild-type *LIG1* cDNA, demonstrating the on-target nature of the genetic tools. Using a degradable DNA Ligase I fusion protein, we demonstrated a strong correlation between DNA Ligase I protein level and viability in *BRCA1* mutant cells. Enzymatically inactive DNA Ligase I mutant protein (*LIG1*<sup>K568A</sup>) was unable to rescue the loss of viability caused by endogenous *LIG1* depletion supporting the tractability of this target from a small molecule inhibitor perspective. These data were reproduced in vivo using *BRCA1* mutant MDA-MB-436 derived tumors, in which tumor growth was inhibited >80% upon loss of *LIG1*.

Mechanistically, DNA ligase I plays a critical role in DNA replication and damage repair by sealing nicks in the phosphodiester backbone of DNA. When these nicks are not repaired, they are marked by the addition of PAR chains. Consistent with this mechanism, we demonstrated that inactivation of *LIG1* leads to increased PARylation. The induction of PARylation was proportional to the level of inactivation of *LIG1* protein, supporting a correlation between *LIG1* activity and DNA nick repair. Together, these data support DNA ligase I as a synthetic lethal target in the context of *BRCA1* mutations.

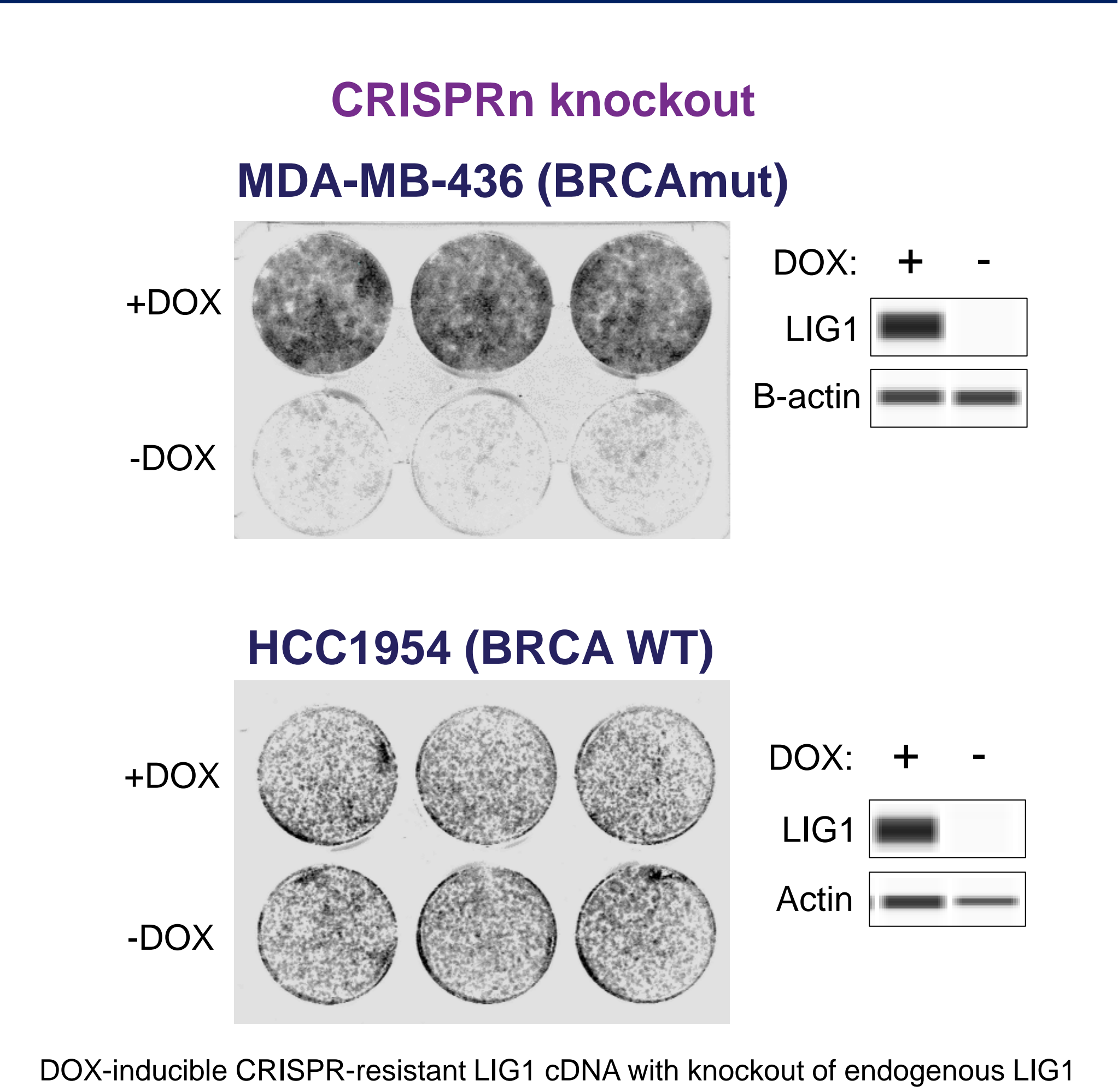
## CRISPR screens identify LIG1 as a top dependency in BRCA mutant cell lines



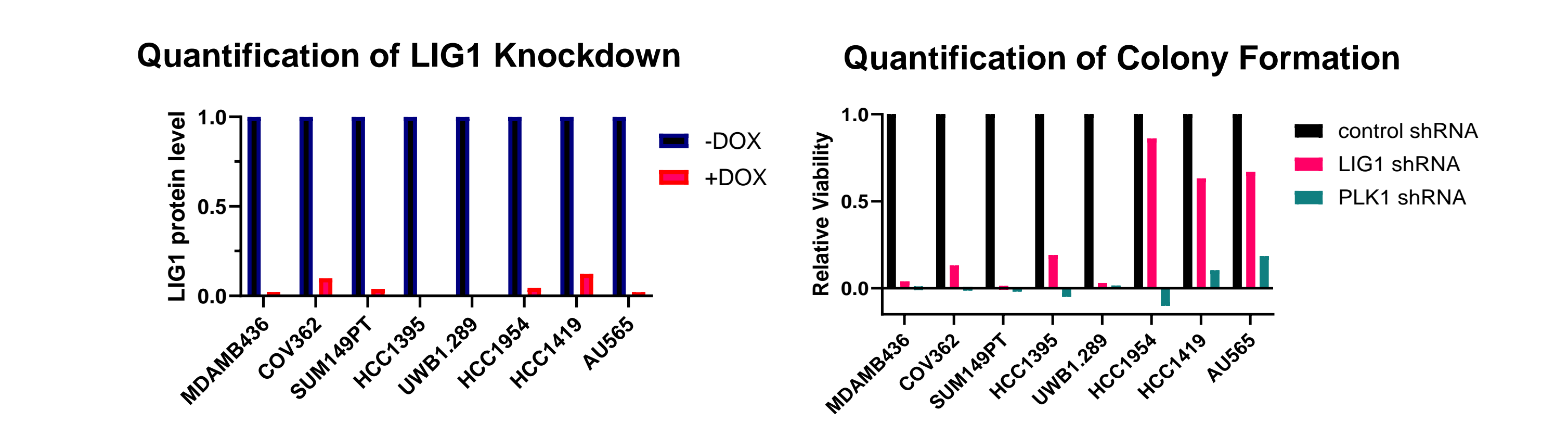
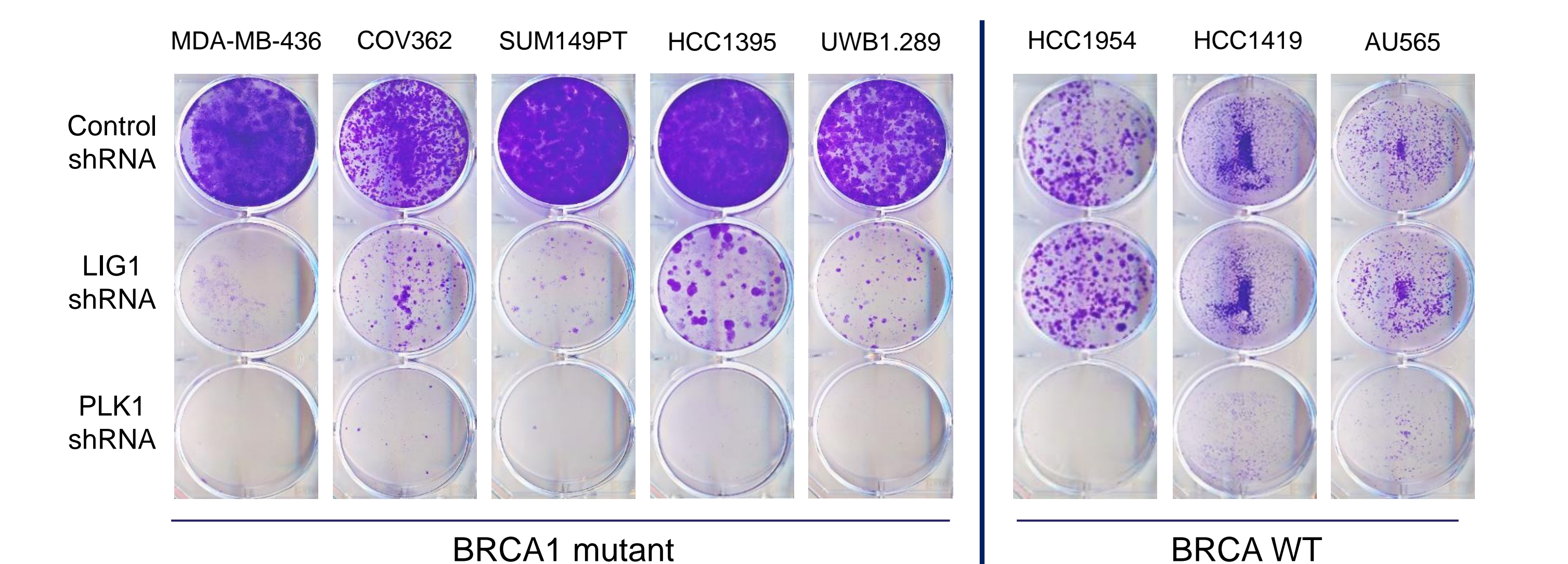
## LIG1 inactivation is lethal in BRCA mutant cells across multiple perturbation platforms



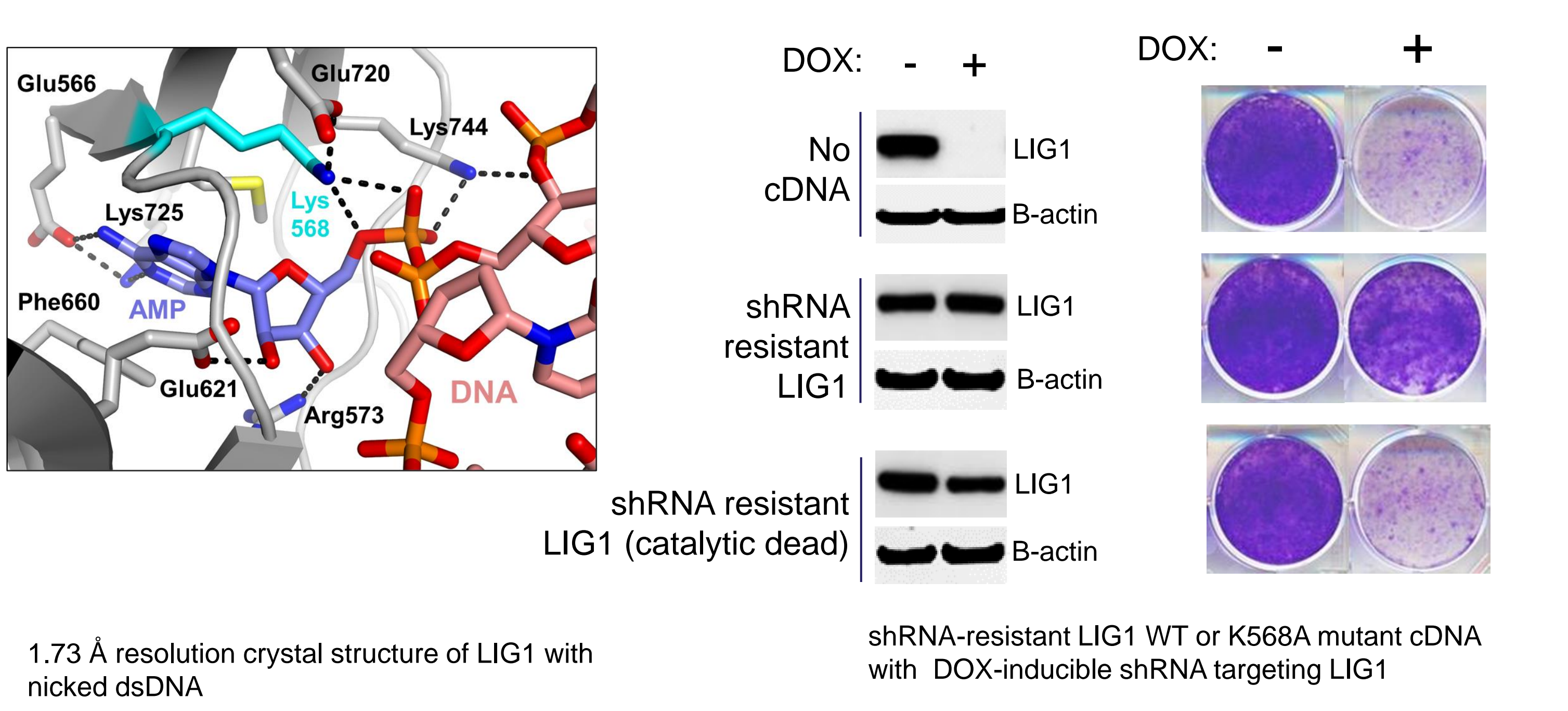
## LIG1 inactivation represses BRCA mutant cell growth in an on-target and selective manner



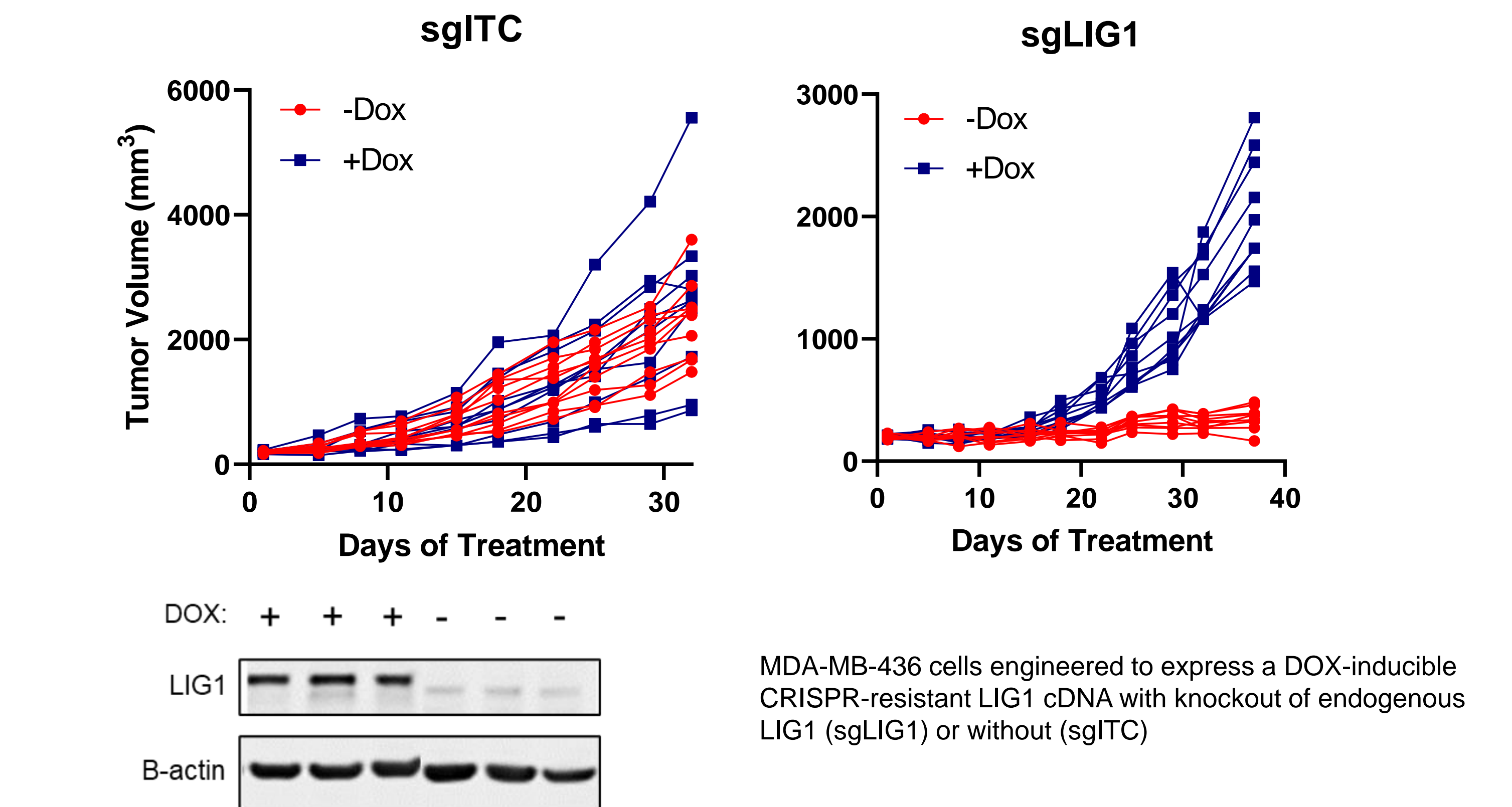
## LIG1 inactivation represses colony formation of BRCA mutant, but not BRCA WT cells



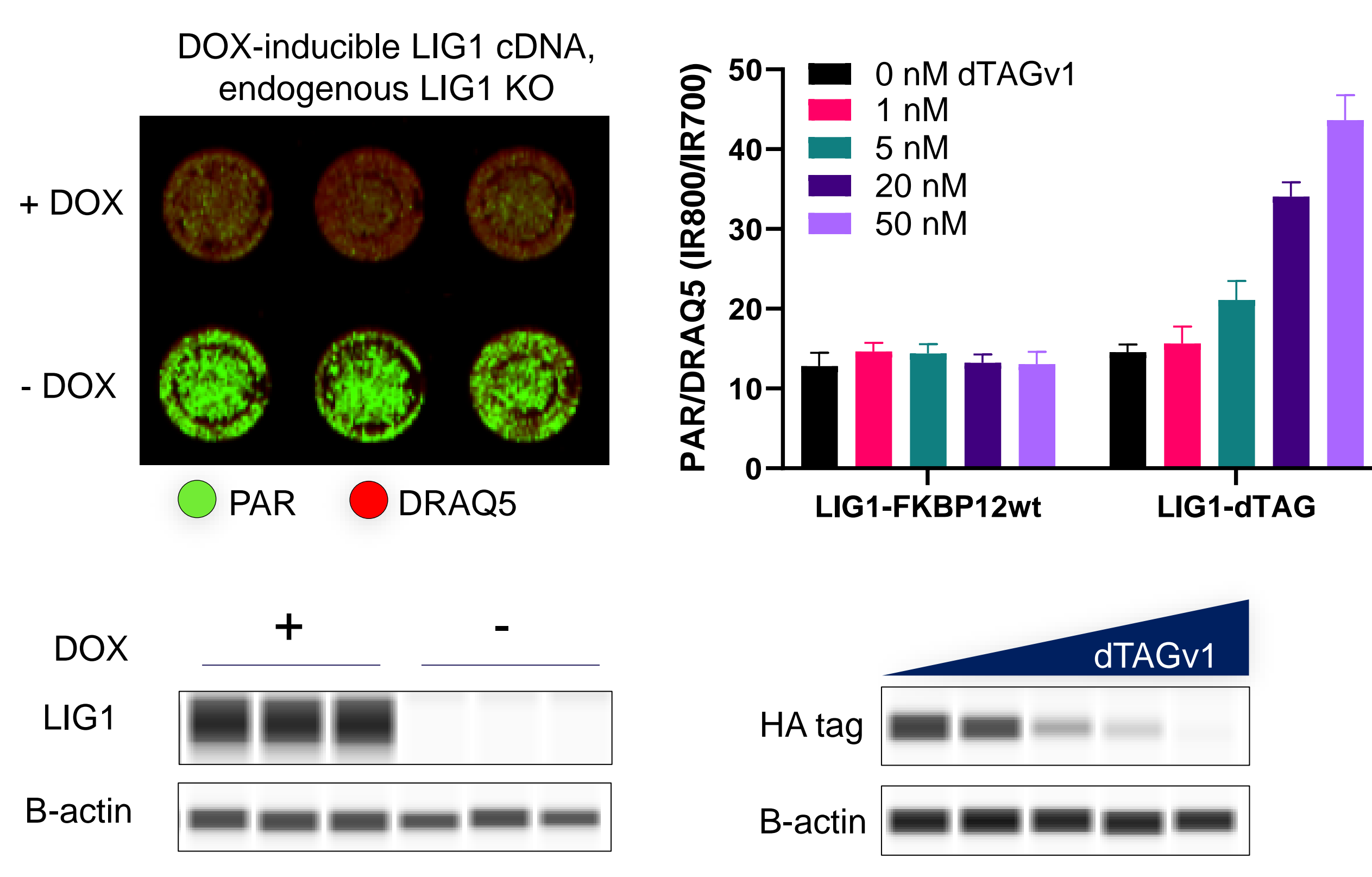
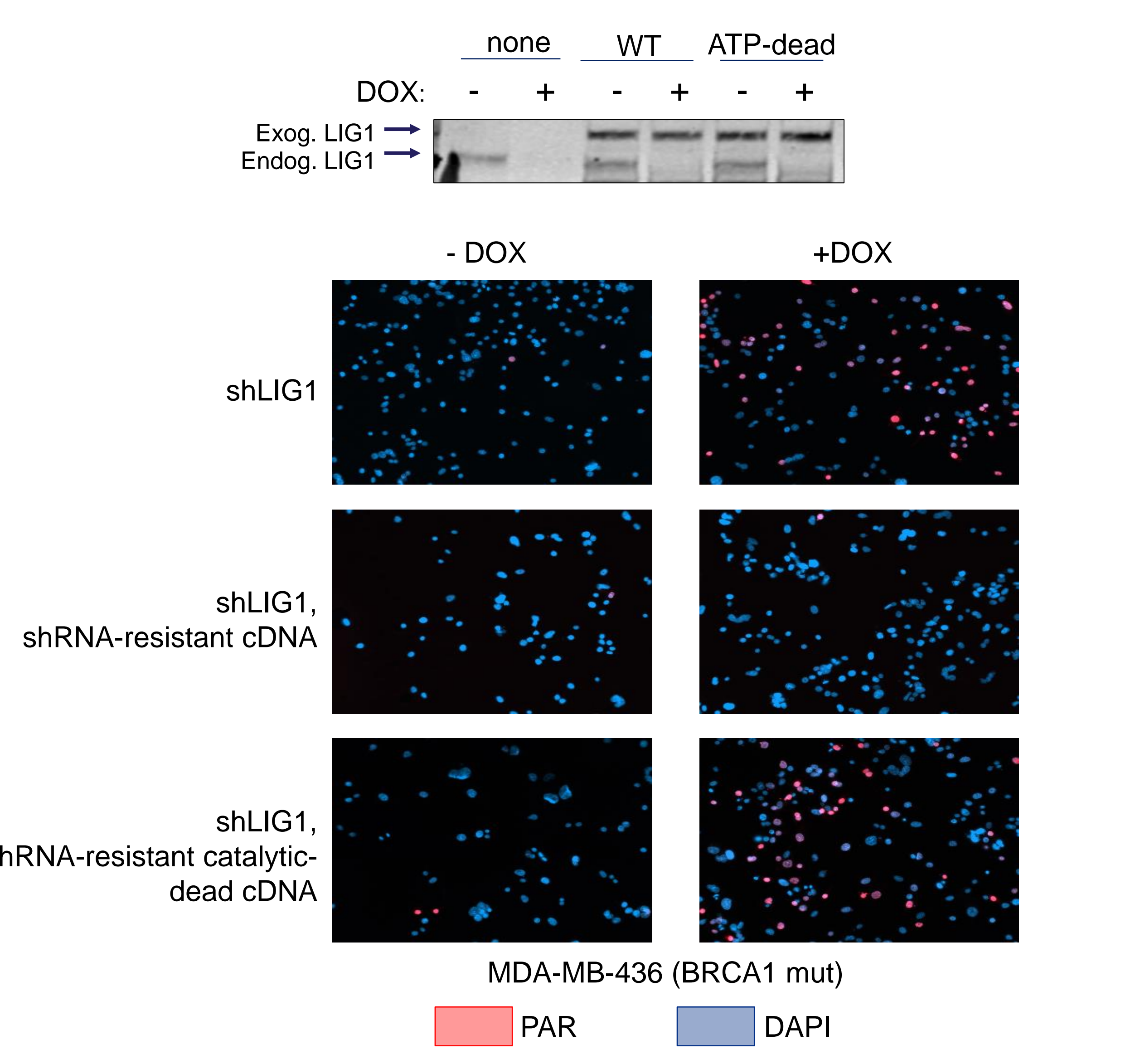
## Catalytic activity of LIG1 is required for viability in BRCA mutant cells



## LIG1 inactivation inhibits BRCA mutant tumor growth in vivo



## Inactivation of LIG1 causes accumulation of PAR chains at DNA strand breaks



## Summary

- Inactivation of *LIG1* is synthetic lethal with *BRCA1* loss in a variety of cell lines
- Inactivation of *LIG1* does not significantly impair viability of *BRCA* WT cells
- Defects in cell viability resultant from *LIG1* inactivation in *BRCA1* mutant cells are on-target and dependent on the catalytic activity of *LIG1*
- LIG1* loss impairs growth of *BRCA1* mutant xenografts resulting in tumor stasis
- Loss of *LIG1* leads to increased PARylation in cells, a readout of accumulation of DNA nicks